

Advancements and Challenges in Biomarkers for Colorectal Cancer Detection: A Comprehensive Review

Yasir Hameed*

Department of Biotechnology, Institute of Biochemistry, Biotechnology, and Bioinformatics, The Islamia University of Bahawalpur, Pakistan

*Corresponding author: Yasir Hameed, yasirhameed2011@gmail.com

Copyright: © 2024 Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), permitting distribution and reproduction in any medium, provided the original work is cited.

Abstract: This study provides an overview of the current landscape of biomarkers for colorectal cancer (CRC) detection, focusing on genetic, proteomic, circulating microRNA (miRNA), and metabolomic biomarkers. CRC remains a significant global health challenge, ranking among the most prevalent cancers worldwide and being a leading cause of cancer-related deaths. Despite advancements in screening methods such as colonoscopy, sigmoidoscopy, and fecal occult blood tests (FOBT), the asymptomatic nature of early-stage CRC often results in late diagnoses, negatively impacting patient outcomes. Genetic biomarkers like *APC*, *KRAS*, *TP53*, and microsatellite instability (MSI) play critical roles in CRC pathogenesis and progression. These biomarkers, detectable through polymerase chain reaction, next-generation sequencing, and other advanced techniques, guide early detection and personalized treatment decisions. Proteomic biomarkers such as CEA, CA 19-9, and novel signatures offer insights into CRC's physiological changes and disease status, aiding prognosis and treatment response assessments through enzyme-linked immunosorbent assay and mass spectrometry. Circulating miRNAs, including *miR-21* and *miR-92a*, present promising non-invasive biomarkers that can be detected in blood and stool samples, reflecting CRC presence, progression, and therapeutic response. Metabolomic biomarkers, encompassing amino acids, lipids, and TCA cycle intermediates, provide further insights into CRC-associated metabolic alterations, which are crucial for early detection and treatment monitoring using mass spectrometry and nuclear magnetic resonance. Despite these advancements, challenges such as biomarker validation, standardization, and clinical utility remain. Future research directions include integrating multi-omics approaches and leveraging technologies like liquid biopsies and AI for enhanced biomarker discovery and clinical application. By addressing these challenges and advancing research in biomarker development, CRC screening and management could potentially be revolutionized, improving patient outcomes and reducing the global burden of this disease.

Keywords: Colorectal cancer; Biomarker; Diagnosis; Detection

Online publication: September 25, 2024

1. Introduction

Colorectal cancer (CRC) is one of the most significant global health challenges, ranking as the third most common cancer and the second leading cause of cancer-related deaths. The burden of CRC is substantial, with over 1.9 million new cases and approximately 935,000 deaths estimated worldwide in 2020 ^[1]. The incidence of CRC varies by geographic region, reflecting differences in risk factors such as diet, lifestyle, and access to healthcare services ^[2].

Early detection of CRC is crucial for improving patient outcomes, as the prognosis is significantly better when the disease is diagnosed at an early stage. The five-year survival rate for localized CRC exceeds 90% but drops to less than 15% for metastatic cases ^[3]. Despite the clear benefits of early detection, many CRC cases are diagnosed at advanced stages due to the asymptomatic nature of early-stage disease and limitations in current screening methods ^[4]. Traditional screening techniques, including colonoscopy, sigmoidoscopy, and fecal occult blood tests (FOBT), such as fecal immunochemical tests (FIT), are commonly used. While colonoscopy is considered the gold standard for CRC detection and prevention, it is invasive, expensive, and often associated with patient discomfort and low compliance rates ^[5]. FOBT and FIT, though non-invasive and more cost-effective, have variable sensitivity and specificity, leading to false positives and negatives ^[6].

In recent years, there has been growing interest in developing biomarkers for CRC detection. Biomarkers are biological molecules found in blood, body fluids, or tissues that indicate a normal or abnormal process, or the presence of a disease ^[7]. They offer the potential for non-invasive, accurate, and early detection of CRC, thereby improving patient compliance and screening efficacy ^[8].

The term “biomarker” encompasses a wide range of biological entities, including genetic mutations, epigenetic alterations, protein expression patterns, metabolites, and circulating tumor components. These biomarkers can be detected in various biological samples such as blood, stool, urine, and tissue biopsies ^[9]. The ideal biomarker for CRC detection should have high sensitivity and specificity, be easily accessible, cost-effective, and applicable across diverse populations ^[10].

This review aims to provide a comprehensive update on the current state of biomarkers for CRC detection. It will explore various types of biomarkers, including genetic, epigenetic, proteomic, metabolomic, and circulating biomarkers, and discuss their roles in early detection and diagnosis. Furthermore, the review will examine the clinical utility of these biomarkers, the challenges associated with their implementation, and future perspectives in the field.

By offering an overview of the latest advancements and ongoing research in CRC biomarkers, this review seeks to highlight the potential of these novel approaches to transform CRC screening and improve patient outcomes. Understanding the current landscape of CRC biomarkers is essential for researchers, clinicians, and policymakers aiming to reduce the global burden of CRC through innovative and effective screening strategies ^[11].

2. Genetic biomarkers

Genetic biomarkers are pivotal for understanding the molecular underpinnings of CRC and play a significant role in early detection and personalized treatment strategies. CRC arises from the accumulation of genetic mutations that drive the transformation of normal colonic epithelium into adenocarcinoma ^[12]. Several key genetic biomarkers have been identified, each contributing to different stages of CRC development and progression. This section delves into the primary genetic biomarkers associated with CRC, including *APC*, *KRAS*, *TP53*, and microsatellite instability (MSI), discussing their roles, detection methods, and clinical implications ^[13-16].

2.1. APC mutations

The adenomatous polyposis coli (*APC*) gene is a tumor suppressor that plays a crucial role in the Wnt signaling pathway. Mutations in *APC* are considered one of the earliest events in colorectal tumorigenesis and are present in approximately 80% of sporadic CRC cases^[17]. These mutations lead to the loss of APC protein function, resulting in the accumulation of β -catenin in the nucleus, where it activates transcription of genes involved in cell proliferation and survival^[18,19].

2.2. KRAS mutations

The *KRAS* gene, encoding a GTPase involved in the EGFR signaling pathway, is another critical genetic biomarker in CRC. Mutations in *KRAS* occur in approximately 35%–45% of CRC cases, typically in codons 12 and 13. These mutations lead to the constitutive activation of the KRAS protein, promoting cell proliferation, survival, and metastasis^[20,21].

2.3. TP53 mutations

The *TP53* gene, which encodes the tumor suppressor protein p53, is often referred to as the “guardian of the genome” due to its role in maintaining genomic stability. *TP53* mutations are present in approximately 50% of CRC cases and often occur at later stages of tumorigenesis, marking the transition from adenoma to carcinoma^[22,23].

2.4. Microsatellite instability

MSI is a form of genomic instability resulting from defects in the DNA mismatch repair (MMR) system. MSI is characterized by the accumulation of insertion or deletion errors in microsatellite regions of the genome. Approximately 15% of CRC cases exhibit high-level MSI (MSI-H), which is commonly associated with Lynch syndrome (hereditary nonpolyposis CRC, HNPCC) and some sporadic CRC cases^[24,25].

2.5. Emerging genetic biomarkers

In addition to well-established genetic biomarkers, ongoing research continues to identify novel genetic alterations with potential clinical utility in CRC detection and management. Emerging biomarkers include:

- (1) *BRAF* mutations: *BRAF* V600E mutations occur in approximately 10% of CRC cases and are associated with poor prognosis and resistance to certain targeted therapies. Detection of *BRAF* mutations can guide treatment decisions and identify patients who may benefit from BRAF inhibitors combined with other therapeutic agents^[26,27].
- (2) *PIK3CA* mutations: Mutations in the *PIK3CA* gene, encoding a subunit of the PI3K enzyme, are found in about 15%–20% of CRC cases. These mutations drive tumor growth and resistance to therapies, making them valuable targets for novel therapeutic strategies^[28,29].
- (3) *NRAS* mutations: *NRAS* mutations, though less common than *KRAS* mutations, are present in a subset of CRC cases. Like *KRAS*, *NRAS* mutations can impact response to anti-EGFR therapies, underscoring the importance of comprehensive RAS testing in CRC^[30,31].

2.6. Detection methods and clinical implications

The detection of genetic biomarkers in CRC utilizes a variety of advanced techniques to ensure accurate identification and subsequent clinical action. *APC* mutations, early events in CRC tumorigenesis, can be detected through polymerase chain reaction (PCR), next-generation sequencing (NGS), and digital droplet PCR (ddPCR), with non-invasive stool DNA testing, such as the Cologuard test, offering a practical option^[32,33].

KRAS mutations, present in about 35%–45% of CRC cases, are identified using PCR-based methods, NGS, and allele-specific oligonucleotide PCR, with liquid biopsy offering a minimally invasive alternative ^[34,35]. *TP53* mutations, found in roughly 50% of CRC cases, are detected through Sanger sequencing, NGS, immunohistochemistry (IHC), and circulating tumor DNA (ctDNA) analysis ^[22,36]. MSI, associated with defects in the MMR system, is assessed using PCR to analyze microsatellite markers and IHC for MMR protein expression, with NGS panels providing comprehensive profiling ^[25,37,38].

Clinically, these biomarkers are crucial for guiding CRC management and treatment. *APC* mutation identification aids in early detection, particularly in individuals with familial adenomatous polyposis (FAP), enabling preventive measures ^[32,33]. *KRAS* mutation status is critical for determining eligibility for anti-EGFR therapies, such as cetuximab and panitumumab, as patients with these mutations do not benefit from such treatments ^[34,35]. The presence of *TP53* mutations, often associated with aggressive disease and poorer prognosis, informs the need for therapies targeting DNA repair pathways or reactivating mutant p53 ^[22,36]. MSI testing, essential for all CRC patients, identifies those likely to respond to immune checkpoint inhibitors and helps in screening for Lynch syndrome ^[25,37]. Emerging biomarkers like *BRAF* and *PIK3CA* mutations also provide insights into therapy resistance and potential targets for novel treatments, while *NRAS* mutations underscore the importance of comprehensive RAS testing to optimize anti-EGFR therapy decisions ^[25,30].

3. Proteomic biomarkers

Proteomic biomarkers, which involve studying the complete set of proteins expressed by a genome, cell, tissue, or organism, hold great promise for the detection and management of CRC. Unlike genetic biomarkers, which provide information about the potential for cancer development, proteomic biomarkers reflect real-time physiological changes and disease states. The dynamic nature of the proteome makes it a rich source for identifying disease-specific alterations that can aid in early detection, prognostication, and therapeutic targeting ^[39,40].

This section explores key proteomic biomarkers in CRC, including carcinoembryonic antigen (CEA), cancer antigen 19-9 (CA 19-9), and novel proteomic signatures, highlighting their detection methods and clinical implications ^[41-43].

3.1. Carcinoembryonic antigen

CEA is one of the most extensively studied and widely used proteomic biomarkers in CRC. CEA is a glycoprotein involved in cell adhesion, and its expression is significantly elevated in colorectal tumors compared to normal tissues ^[41,44].

3.2. Cancer antigen 19-9

CA 19-9 is another glycoprotein that serves as a tumor marker in various gastrointestinal cancers, including CRC. Although CA 19-9 is more commonly associated with pancreatic cancer, it can also be elevated in a subset of CRC patients ^[41,45].

3.3. Novel proteomic signatures

Advances in proteomics technologies, such as mass spectrometry and protein microarrays, have facilitated the discovery of novel proteomic signatures that can improve CRC detection and management. These signatures often comprise multiple protein biomarkers that together enhance diagnostic accuracy and provide insights into the molecular mechanisms of CRC ^[42,43].

3.4. Circulating tumor proteins

Circulating tumor proteins, released by tumor cells into the bloodstream, represent another important category of proteomic biomarkers. These proteins can be detected in blood samples, providing a minimally invasive approach to CRC detection and monitoring [42,43].

3.5. Detection methods and clinical implications

Proteomic biomarkers for CRC, such as CEA, CA 19-9, novel proteomic signatures, and circulating tumor proteins, are detected using advanced techniques and have significant clinical implications. CEA and CA 19-9 levels are typically measured using enzyme-linked immunosorbent assays (ELISAs) or chemiluminescent immunoassays (CLIAs) in serum samples, providing a non-invasive method for monitoring disease progression and treatment response [41,44]. While CEA is valuable for postoperative monitoring and detecting recurrence, CA 19-9, though less specific to CRC, can complement CEA in assessing disease status [45].

Novel proteomic signatures, identified through high-throughput platforms like mass spectrometry (MS) and protein microarrays, offer comprehensive profiling that enhances diagnostic accuracy and provides insights into CRC's molecular mechanisms [42,43]. These signatures are crucial for early detection, patient stratification, and identifying therapeutic targets. Circulating tumor proteins, detected through techniques like ELISA, bead-based multiplex assays, and MS, provide a minimally invasive approach for early detection and monitoring. Elevated levels of these proteins in blood samples can indicate tumor presence, progression, and treatment response, aiding in timely CRC management [42,43].

Integrating these proteomic biomarkers into clinical practice enhances early detection, informs prognosis, guides personalized therapy, and ultimately improves patient outcomes.

4. Circulating microRNAs

Circulating microRNAs (miRNAs) have emerged as promising non-invasive biomarkers for CRC detection, prognosis, and monitoring. These small, non-coding RNA molecules, typically 19–25 nucleotides in length, regulate gene expression post-transcriptionally and can be found in various body fluids, including blood, serum, plasma, and stool [46,47]. The stability of miRNAs in circulation, owing to their protection within exosomes, microvesicles, or protein complexes, makes them attractive candidates for clinical applications. This section explores key circulating miRNAs, their detection methods, and their clinical implications in CRC.

4.1. Key circulating miRNAs

- (1) *miR-21*: One of the most extensively studied miRNAs in CRC, *miR-21* is frequently overexpressed in CRC tissues and detectable at elevated levels in the blood of CRC patients [48,49].
- (2) *miR-92a*: Part of the *miR-17-92* cluster, *miR-92a* is upregulated in CRC and is associated with tumor growth and metastasis [50,51].
- (3) *miR-29a*: This miRNA is involved in the regulation of apoptosis and cell proliferation, with increased levels observed in the blood of CRC patients [52,53].
- (4) *miR-17-3p* and *miR-20a*: Both are part of the *miR-17-92* cluster and are implicated in CRC progression and metastasis [54,55].
- (5) *miR-145*: Generally downregulated in CRC, *miR-145* acts as a tumor suppressor, and its reduced expression correlates with more advanced disease stages [56,57].

4.2. Detection methods and clinical implications

Circulating miRNAs are detected using techniques such as quantitative PCR (qPCR), ddPCR, NGS, microarrays, and bead-based multiplex assays. These methods enable the sensitive and specific quantification of miRNAs in body fluids [58,59]. Clinically, circulating miRNAs hold significant potential for early CRC detection, prognosis, and monitoring of treatment response. Elevated levels of miRNAs, such as *miR-21*, *miR-92a*, and *miR-29a*, in blood samples can indicate the presence of CRC, even at early stages, facilitating timely intervention [53,60]. Furthermore, the expression profiles of specific miRNAs provide prognostic information, correlating with disease progression and patient outcomes [49,61]. Changes in miRNA levels during and after treatment can reflect therapeutic efficacy and help detect recurrence, aiding in the development of personalized treatment strategies.

The non-invasive nature of circulating miRNA testing offers a patient-friendly alternative to traditional biopsies, making routine monitoring more accessible and less burdensome. Integrating miRNA profiles into clinical practice enhances the precision of CRC diagnosis, prognosis, and treatment, ultimately improving patient outcomes through personalized medicine.

5. Metabolomic biomarkers

Metabolomics, the comprehensive study of metabolites in biological systems, has become an invaluable approach for identifying biomarkers in CRC [62]. Metabolites are small molecules involved in various metabolic pathways, reflecting the physiological state of cells, tissues, and organisms [63]. Changes in metabolite levels can indicate alterations in metabolic processes associated with cancer development and progression. This section explores key metabolomic biomarkers in CRC, their detection methods, and their clinical implications.

5.1. Key metabolomic biomarkers

- (1) Amino acids: Altered levels of amino acids, such as tryptophan, glutamine, and arginine, have been observed in CRC patients. These changes reflect disruptions in amino acid metabolism, which is crucial for tumor growth and survival [64,65].
- (2) Lipid metabolites: Abnormal lipid metabolism is a hallmark of cancer. Elevated levels of certain phospholipids, sphingolipids, and free fatty acids are commonly found in CRC patients [66,67].
- (3) Carbohydrate metabolites: Changes in carbohydrate metabolism, including elevated levels of glucose and lactate, are indicative of the Warburg effect—a phenomenon where cancer cells preferentially utilize glycolysis for energy production even in the presence of oxygen [68,69].
- (4) Bile acids: Altered bile acid profiles have been linked to CRC. Elevated levels of primary and secondary bile acids can reflect changes in gut microbiota and hepatic function [70,71].
- (5) Tricarboxylic acid cycle intermediates: Disruptions in the tricarboxylic acid (TCA) cycle, such as altered levels of citrate, succinate, and fumarate, indicate metabolic reprogramming in cancer cells [72,73].

5.2. Detection methods and clinical implications

Metabolomic biomarkers in CRC are detected using advanced analytical techniques, such as MS, nuclear magnetic resonance (NMR) spectroscopy, capillary electrophoresis-mass spectrometry (CE-MS), Fourier transform infrared (FTIR) spectroscopy, and high-performance liquid chromatography (HPLC) [63,65]. These methods enable the sensitive and specific identification and quantification of metabolites in biological samples, providing a detailed metabolic profile of both the tumor and the host. Clinically, metabolomic biomarkers

have significant implications for CRC management. They facilitate early detection through the identification of specific metabolic changes associated with cancer onset ^[64,69]. Certain metabolites, such as altered amino acids, lipids, carbohydrates, bile acids, and TCA cycle intermediates, offer prognostic value by correlating with disease stage, progression, and patient outcomes ^[70,72]. Metabolomic biomarkers are also valuable for monitoring treatment response and detecting recurrence, as changes in metabolite levels can reflect therapeutic efficacy and disease status ^[66,73]. Furthermore, metabolomic profiling can guide personalized therapy by identifying metabolic vulnerabilities that can be targeted with specific treatments ^[65,68]. The non-invasive nature of metabolomic testing, using fluids such as blood, urine, and stool, enhances patient comfort and facilitates routine monitoring. Integrating metabolomic data with other omics data provides a comprehensive understanding of CRC, leading to improved disease management and patient outcomes through personalized medicine.

6. Challenges and future directions

Biomarker research for CRC faces several significant challenges that must be addressed to realize its full potential in clinical practice. One critical challenge is the standardization and validation of biomarkers. Biomarker discovery often involves diverse methodologies and sample types across different studies, leading to variability in results. Standardizing protocols for biomarker identification, validation, and clinical implementation is essential to ensure reproducibility and reliability across various research settings and populations.

Another key hurdle is demonstrating the clinical utility of biomarkers and integrating them into routine clinical practice. While biomarkers show promise in research, their adoption in clinical settings requires robust evidence of their effectiveness in improving patient outcomes. Biomarkers must demonstrate clear benefits in terms of sensitivity, specificity, cost-effectiveness, and impact on clinical decision-making to justify their incorporation into screening and diagnostic algorithms.

The heterogeneity of CRC presents another challenge. CRC includes various molecular subtypes and clinical manifestations, requiring biomarkers that can accurately reflect this diversity. Biomarkers must be validated across different patient populations to ensure their efficacy in stratifying patients for personalized treatment strategies.

Ethnic and geographic variations also influence biomarker performance. Genetic, lifestyle, and environmental factors can affect biomarker expression and efficacy across different ethnic groups and geographic regions. Developing biomarkers that are effective and reliable across diverse populations is crucial for their global applicability and adoption in clinical practice.

Additionally, the transition from single biomarkers to multimodal biomarker panels represents a promising future direction in CRC research. Single biomarkers may lack sufficient sensitivity or specificity for accurate CRC detection and prognosis. Combining multiple types of biomarkers, such as genetic, proteomic, and metabolomic markers, could enhance diagnostic accuracy and reliability, paving the way for more effective screening and personalized treatment approaches.

Looking ahead, advancements in multi-omics approaches—including genomics, epigenomics, proteomics, and metabolomics—hold great potential for improving biomarker discovery and validation. Integrating data from multiple omics layers can provide a more comprehensive understanding of CRC biology and facilitate the development of robust biomarker panels. Moreover, leveraging technologies such as liquid biopsies and artificial intelligence (AI) for biomarker detection and analysis could further enhance the clinical utility and predictive power of biomarkers in CRC management.

Addressing these challenges and pursuing these future directions will be instrumental in advancing biomarker research for CRC. By overcoming these obstacles, biomarkers have the potential to revolutionize CRC screening, diagnosis, and treatment, ultimately improving patient outcomes and reducing the global burden of this disease.

7. Conclusion

Biomarkers present a promising pathway for the early detection and management of CRC. Genetic, proteomic, circulating miRNA, and metabolomic biomarkers have demonstrated potential in enhancing screening accuracy and improve patient outcomes. However, significant challenges remain in the standardization, validation, and clinical integration of these biomarkers. Future research should focus on developing robust, multimodal biomarker panels and utilizing technological advancements to strengthen CRC detection and screening programs. With continued efforts, biomarker-based screening could become a cornerstone in the fight against CRC, ultimately reducing its global impact.

Disclosure statement

The authors declare no conflict of interest.

References

- [1] Sung H, Ferlay J, Siegel RL, et al., 2021, Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin*, 71(3): 209–249. <https://doi.org/10.3322/caac.21660>
- [2] Siegel RL, Miller KD, Jemal A, 2019, Cancer Statistics, 2019. *CA Cancer J Clin*, 69(1): 7–34. <https://doi.org/10.3322/caac.21551>
- [3] Nguyen HT, Duong HQ, 2018, The Molecular Characteristics of Colorectal Cancer: Implications for Diagnosis and Therapy. *Oncol Lett*, 16(1): 9–18. <https://doi.org/10.3892/ol.2018.8679>
- [4] Guinney J, Dienstmann R, Wang X, et al., 2015, The Consensus Molecular Subtypes of Colorectal Cancer. *Nat Med*, 21(11): 1350–1356. <https://doi.org/10.1038/nm.3967>
- [5] Sveen A, Kopetz S, Lothe RA, 2020, Biomarker-Guided Therapy for Colorectal Cancer: Strength in Complexity. *Nat Rev Clin Oncol*, 17(1): 11–32. <https://doi.org/10.1038/s41571-019-0241-1>
- [6] Imperiale TF, 2012, Noninvasive Screening Tests for Colorectal Cancer. *Dig Dis*, 30(2): 16–26. <https://doi.org/10.1159/000341884>
- [7] Hanash SM, Pitteri SJ, Faca VM, 2008, Mining the Plasma Proteome for Cancer Biomarkers. *Nature*, 452(7187): 571–579. <https://doi.org/10.1038/nature06916>
- [8] Pizzini S, Bisognin A, Mandruzzato S, et al., 2013, Impact of MicroRNAs on Regulatory Networks and Pathways in Human Colorectal Carcinogenesis and Development of Metastasis. *BMC Genomics*, 14: 589. <https://doi.org/10.1186/1471-2164-14-589>
- [9] Mäbert K, Cojoc M, Peitzsch C, et al., 2014, Cancer Biomarker Discovery: Current Status and Future Perspectives. *Int J Radiat Biol*, 90(8): 659–677. <https://doi.org/10.3109/09553002.2014.892229>
- [10] Das V, Kalita J, Pal M, 2017, Predictive and Prognostic Biomarkers in Colorectal Cancer: A Systematic Review of Recent Advances and Challenges. *Biomed Pharmacother*, 87: 8–19. <https://doi.org/10.1016/j.biopha.2016.12.064>
- [11] Bresalier RS, Senore C, Young GP, et al., 2023, An Efficient Strategy for Evaluating New Non-Invasive Screening Tests

for Colorectal Cancer: The Guiding Principles. *Gut*, 72(10): 1904–1918. <https://doi.org/10.1136/gutjnl-2023-329701>

- [12] Pellino G, Gallo G, Pallante P, et al., 2018, Noninvasive Biomarkers of Colorectal Cancer: Role in Diagnosis and Personalised Treatment Perspectives. *Gastroenterol Res Pract*, 2018: 2397863. <https://doi.org/10.1155/2018/2397863>
- [13] Vargas AC, McCart Reed AE, Waddell N, et al., 2012, Gene Expression Profiling of Tumour Epithelial and Stromal Compartments During Breast Cancer Progression. *Breast Cancer Res Treat*, 135(1): 153–165. <https://doi.org/10.1007/s10549-012-2123-4>
- [14] Zheng E, Włodarczyk M, Węgiel A, et al., 2024, Navigating Through Novelties Concerning mCRC Treatment – The Role of Immunotherapy, Chemotherapy, and Targeted Therapy in mCRC. *Front Surg*, 11: 1398289. <https://doi.org/10.3389/fsurg.2024.1398289>
- [15] Saadh MJ, Allela OQB, Sattay ZJ, et al., 2024, Deciphering the Functional Landscape and Therapeutic Implications of Noncoding RNAs in the TGF- β Signaling Pathway in Colorectal Cancer: A Comprehensive Review. *Pathol Res Pract*, 255: 155158.
- [16] Lee SY, Haq F, Kim D, et al., 2014, Comparative Genomic Analysis of Primary and Synchronous Metastatic Colorectal Cancers. *PLoS One*, 9(3): e90459. <https://doi.org/10.1371/journal.pone.0090459>. Erratum in *PLoS One*, 10(1): e0117753. <https://doi.org/10.1371/journal.pone.0117753>
- [17] Smith G, Carey FA, Beattie J, et al., 2002, Mutations in APC, Kirsten-ras, and p53–Alternative Genetic Pathways to Colorectal Cancer. *Proc Natl Acad Sci U S A*, 99(14): 9433–9438. <https://doi.org/10.1073/pnas.122612899>
- [18] Jones S, Chen WD, Parmigiani G, et al., 2008, Comparative Lesion Sequencing Provides Insights into Tumor Evolution. *Proc Natl Acad Sci U S A*, 105(11): 4283–4288. <https://doi.org/10.1073/pnas.0712345105>
- [19] Thewjitcharoen Y, Shuangshoti S, Lerdlum S, et al., 2014, Colorectal Cancer Manifesting with Metastasis to Prolactinoma: Report of A Case Involving Symptoms Mimicking Pituitary Apoplexy. *Intern Med*, 53(17): 1965–1969. <https://doi.org/10.2169/internalmedicine.53.2353>
- [20] Guan C, Zhang X, Yu L, 2024, A Review of Recent Advances in the Molecular Mechanisms Underlying Brain Metastasis in Lung Cancer. *Mol Cancer Ther*, 23(5): 627–637. <https://doi.org/10.1158/1535-7163.MCT-23-0416>
- [21] Martinelli E, Troiani T, Sforza V, et al., 2018, Sequential HER2 Blockade as Effective Therapy in Chemorefractory, HER2 Gene-Amplified, RAS Wild-Type, Metastatic Colorectal Cancer: Learning From A Clinical Case. *ESMO Open*, 3(1): e000299. <https://doi.org/10.1136/esmoopen-2017-000299>. Erratum in *ESMO Open*, 4(2): e000299corr1. <https://doi.org/10.1136/esmoopen-2017-000299corr1>
- [22] Cancer Genome Atlas Network, 2012, Comprehensive Molecular Characterization of Human Colon and Rectal Cancer. *Nature*, 487(7407): 330–337. <https://doi.org/10.1038/nature11252>
- [23] Kandoth C, McLellan MD, Vandin F, et al., 2013, Mutational Landscape and Significance Across 12 Major Cancer Types. *Nature*, 502(7471): 333–339. <https://doi.org/10.1038/nature12634>
- [24] Boland CR, Goel A, 2010, Microsatellite Instability in Colorectal Cancer. *Gastroenterology*, 138(6): 2073–2087.e3. <https://doi.org/10.1053/j.gastro.2009.12.064>
- [25] Le DT, Uram JN, Wang H, et al., 2015, PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N Engl J Med*, 372(26): 2509–2520. <https://doi.org/10.1056/NEJMoa1500596>
- [26] Davies H, Bignell GR, Cox C, et al., 2002, Mutations of the BRAF Gene in Human Cancer. *Nature*, 417(6892): 949–954. <https://doi.org/10.1038/nature00766>
- [27] Kopetz S, Grothey A, Yaeger R, et al., 2019, Encorafenib, Binimetinib, and Cetuximab in BRAF V600E-Mutated Colorectal Cancer. *N Engl J Med*, 381(17): 1632–1643. <https://doi.org/10.1056/NEJMoa1908075>
- [28] Samuels Y, Diaz LA Jr, Schmidt-Kittler O, et al., 2005, Mutant PIK3CA Promotes Cell Growth and Invasion of Human Cancer Cells. *Cancer Cell*, 7(6): 561–573. <https://doi.org/10.1016/j.ccr.2005.05.014>
- [29] Liao X, Morikawa T, Lochhead P, et al., 2012, Prognostic Role of PIK3CA Mutation in Colorectal Cancer: Cohort Study

- and Literature Review. *Clin Cancer Res*, 18(8): 2257–2268. <https://doi.org/10.1158/1078-0432.CCR-11-2410>
- [30] Saridaki Z, Tzardi M, Sfakianaki M, et al., 2013, BRAFV600E Mutation Analysis in Patients with Metastatic Colorectal Cancer (mCRC) in Daily Clinical Practice: Correlations with Clinical Characteristics, and Its Impact on Patients' Outcome. *PLoS One*, 8(12): e84604. <https://doi.org/10.1371/journal.pone.0084604>
- [31] Tran B, Kopetz S, Tie J, et al., 2011, Impact of BRAF Mutation and Microsatellite Instability on the Pattern of Metastatic Spread and Prognosis in Metastatic Colorectal Cancer. *Cancer*, 117(20): 4623–4632. <https://doi.org/10.1002/cncr.26086>
- [32] Imperiale TF, Ransohoff DF, Itzkowitz SH, et al., 2004, Fecal DNA Versus Fecal Occult Blood for Colorectal-Cancer Screening in An Average-Risk Population. *N Engl J Med*, 351(26): 2704–2714. <https://doi.org/10.1056/NEJMoa033403>
- [33] NCCN Clinical Practice Guidelines in Oncology, 2020, Colorectal Cancer Screening, https://www.nccn.org/professionals/physician_gls/pdf/colorectal_screening.pdf
- [34] Amado RG, Wolf M, Peeters M, et al., 2008, Wild-Type KRAS is Required for Panitumumab Efficacy in Patients with Metastatic Colorectal Cancer. *J Clin Oncol*, 26(10): 1626–1634. <https://doi.org/10.1200/JCO.2007.14.7116>
- [35] Diaz A, Park K, Lim DA, et al., 2012, Normalization, Bias Correction, and Peak Calling for ChIP-seq. *Stat Appl Genet Mol Biol*, 11(3): Article 9. <https://doi.org/10.1515/1544-6115.1750>
- [36] Donehower LA, Soussi T, Korkut A, et al., 2019, Integrated Analysis of TP53 Gene and Pathway Alterations in The Cancer Genome Atlas. *Cell Rep*, 28(5): 1370–1384.e5. <https://doi.org/10.1016/j.celrep.2019.07.001>. Erratum in *Cell Rep*, 28(11): 3010. <https://doi.org/10.1016/j.celrep.2019.08.061>
- [37] Boland CR, Thibodeau SN, Hamilton SR, et al., 1998, A National Cancer Institute Workshop on Microsatellite Instability for Cancer Detection and Familial Predisposition: Development of International Criteria for the Determination of Microsatellite Instability in Colorectal Cancer. *Cancer Res*, 58(22): 5248–5257.
- [38] Sargent DJ, Marsoni S, Monges G, et al., 2010, Defective Mismatch Repair as A Predictive Marker for Lack of Efficacy of Fluorouracil-Based Adjuvant Therapy in Colon Cancer. *J Clin Oncol*, 28(20): 3219–3226. <https://doi.org/10.1200/JCO.2009.27.1825>. Erratum in *J Clin Oncol*, 28(30): 4664.
- [39] Anderson NL, Anderson NG, 2002, The Human Plasma Proteome: History, Character, and Diagnostic Prospects. *Mol Cell Proteomics*, 1(11): 845–867. <https://doi.org/10.1074/mcp.r200007-mcp200>. Erratum in *Mol Cell Proteomics*, 2(1): 50.
- [40] Domon B, Aebersold R, 2006, Mass Spectrometry and Protein Analysis. *Science*, 312(5771): 212–217. <https://doi.org/10.1126/science.1124619>
- [41] Kumar VR, Kampan NC, Abd Aziz NH, et al., 2023, Recent Advances in Surface Plasmon Resonance (SPR) Technology for Detecting Ovarian Cancer Biomarkers. *Cancers (Basel)*, 15(23): 5607. <https://doi.org/10.3390/cancers15235607>
- [42] Li C, Sun YD, Yu GY, et al., 2020, Integrated Omics of Metastatic Colorectal Cancer. *Cancer Cell*, 38(5): 734–747.e9. <https://doi.org/10.1016/j.ccell.2020.08.002>
- [43] Zhang W, Yang C, Wang S, et al., 2021, SDC2 and TFPI2 Methylation in Stool Samples as an Integrated Biomarker for Early Detection of Colorectal Cancer. *Cancer Manag Res*, 13: 3601–3617. <https://doi.org/10.2147/CMAR.S300861>. Erratum in *Cancer Manag Res*, 14: 1845–1846. <https://doi.org/10.2147/CMAR.S375358>
- [44] Grunnet M, Sorensen JB, 2012, Carcinoembryonic Antigen (CEA) as Tumor Marker in Lung Cancer. *Lung Cancer*, 76(2): 138–143. <https://doi.org/10.1016/j.lungcan.2011.11.012>
- [45] Locker GY, Hamilton S, Harris J, et al., 2006, ASCO 2006 Update of Recommendations for the Use of Tumor Markers in Gastrointestinal Cancer. *J Clin Oncol*, 24(33): 5313–5327. <https://doi.org/10.1200/JCO.2006.08.2644>
- [46] Cortez MA, Bueso-Ramos C, Ferdin J, et al., 2011, MicroRNAs in Body Fluids – The Mix of Hormones and Biomarkers. *Nat Rev Clin Oncol*, 8(8): 467–477. <https://doi.org/10.1038/nrclinonc.2011.76>
- [47] Arman K, Dalloul Z, Bozgeyik E, 2023, Emerging Role of microRNAs and Long Non-Coding RNAs in COVID-19

with Implications to Therapeutics. *Gene*, 861: 147232. <https://doi.org/10.1016/j.gene.2023.147232>

- [48] Liu Q, Yang W, Luo Y, et al., 2018, Correlation between miR-21 and miR-145 and the Incidence and Prognosis of Colorectal Cancer. *J BUON*, 23(1): 29–35.
- [49] Toiyama Y, Hur K, Tanaka K, et al., 2014, Serum miR-200c is A Novel Prognostic and Metastasis-Predictive Biomarker in Patients with Colorectal Cancer. *Ann Surg*, 259(4): 735–43. <https://doi.org/10.1097/SLA.0b013e3182a6909d>
- [50] Volinia S, Calin GA, Liu CG, et al., 2006, A MicroRNA Expression Signature of Human Solid Tumors Defines Cancer Gene Targets. *Proc Natl Acad Sci U S A*, 103(7): 2257–2261. <https://doi.org/10.1073/pnas.0510565103>
- [51] Bandrés E, Cubedo E, Agirre X, et al., 2006, Identification by Real-time PCR of 13 Mature microRNAs Differentially Expressed in Colorectal Cancer and Non-Tumoral Tissues. *Mol Cancer*, 5: 29. <https://doi.org/10.1186/1476-4598-5-29>
- [52] Wang K, Yuan Y, Cho J-H, et al., 2012, Comparing the MicroRNA Spectrum between Serum and Plasma. *PLoS ONE*, 7(7): e41561. <https://doi.org/10.1371/journal.pone.0041561>
- [53] Wang J, Xu J, Fu J, et al., 2021, MiR-29a Regulates Radiosensitivity in Human Intestinal Cells by Targeting PTEN Gene. *Radiat Res*, 186(3): 292–301. <https://doi.org/10.1667/RR14428.1>
- [54] Pídková P, Herichová I, 2021, miRNA Clusters with Up-Regulated Expression in Colorectal Cancer. *Cancers (Basel)*, 13(12): 2979. <https://doi.org/10.3390/cancers13122979>
- [55] Fang L, Li H, Wang L, et al., 2014, MicroRNA-17-5p Promotes Chemotherapeutic Drug Resistance and Tumour Metastasis of Colorectal Cancer by Repressing PTEN Expression. *Oncotarget*, 5(10): 2974–2987. <https://doi.org/10.18632/oncotarget.1614>
- [56] Michael MZ, O'Connor SM, van Holst Pellekaan NG, et al., 2003, Reduced Accumulation of Specific MicroRNAs in Colorectal Neoplasia. *Mol Cancer Res*, 1(12): 882–891.
- [57] Slaby O, Svoboda M, Fabian P, et al., 2008, Altered Expression of miR-21, miR-31, miR-143 and miR-145 is Related to Clinicopathologic Features of Colorectal Cancer. *Oncology*, 72(5–6): 397–402.
- [58] Witwer KW, Buzás EI, Bemis LT, et al., 2013, Standardization of Sample Collection, Isolation and Analysis Methods in Extracellular Vesicle Research. *J Extracell Vesicles*, 2. <https://doi.org/10.3402/jev.v2i0.20360>
- [59] Schwarzenbach H, Nishida N, Calin GA, et al., 2014, Clinical Relevance of Circulating Cell-Free MicroRNAs in Cancer. *Nat Rev Clin Oncol*, 11(3): 145–156. <https://doi.org/10.1038/nrclinonc.2014.5>
- [60] Igder S, Zamani M, Fakher S, et al., 2024, Circulating Nucleic Acids in Colorectal Cancer: Diagnostic and Prognostic Value. *Dis Markers*, 2024: 9943412. <https://doi.org/10.1155/2024/9943412>
- [61] Ng EK, Chong WW, Jin H, et al., 2009, Differential Expression of MicroRNAs in Plasma of Patients with Colorectal Cancer: A Potential Marker for Colorectal Cancer Screening. *Gut*, 58(10): 1375–1381. <https://doi.org/10.1136/gut.2008.167817>
- [62] Johnson CH, Gonzalez FJ, 2012, Challenges and Opportunities of Metabolomics. *J Cell Physiol*, 227(8): 2975–2981. <https://doi.org/10.1002/jcp.24002>
- [63] Beger RD, Dunn W, Schmidt MA, et al., 2016, Metabolomics Enables Precision Medicine: “A White Paper, Community Perspective”. *Metabolomics*, 12(10): 149. <https://doi.org/10.1007/s11306-016-1094-6>
- [64] Mayers JR, Wu C, Clish CB, et al., 2014, Elevation of Circulating Branched-Chain Amino Acids is An Early Event in Human Pancreatic Adenocarcinoma Development. *Nat Med*, 20(10): 1193–1198. <https://doi.org/10.1038/nm.3686>
- [65] Ragni M, Fornelli C, Nisoli E, et al., 2022, Amino Acids in Cancer and Cachexia: An Integrated View. *Cancers (Basel)*, 14(22): 5691. <https://doi.org/10.3390/cancers14225691>
- [66] Hilvo M, Denkert C, Lehtinen L, et al., 2011, Novel Theranostic Opportunities Offered by Characterization of Altered Membrane Lipid Metabolism in Breast Cancer Progression. *Cancer Res*, 71(9): 3236–3245. <https://doi.org/10.1158/0008-5472.CAN-10-3894>
- [67] Scheurlen KM, Billeter AT, O'Brien SJ, et al., 2020, Metabolic Dysfunction and Early-Onset Colorectal Cancer – How

Macrophages Build The Bridge. *Cancer Med*, 9(18): 6679–6693. <https://doi.org/10.1002/cam4.3315>

- [68] Søreide K, Ismail W, Roalsø M, et al., 2023, Early Diagnosis of Pancreatic Cancer: Clinical Premonitions, Timely Precursor Detection and Increased Curative-Intent Surgery. *Cancer Control*, 30: 10732748231154711. <https://doi.org/10.1177/10732748231154711>
- [69] Xu Y, Dong X, Qin C, et al., 2023, Metabolic Biomarkers in Lung Cancer Screening and Early Diagnosis (Review). *Oncol Lett*, 25(6): 265. <https://doi.org/10.3892/ol.2023.13851>
- [70] van Best N, Rolle-Kampeczyk U, Schaap FG, et al., 2020, Bile Acids Drive the Newborn's Gut Microbiota Maturation. *Nat Commun*, 11(1): 3692. <https://doi.org/10.1038/s41467-020-17183-8>
- [71] Kiriya Y, Nochi H, 2021, Physiological Role of Bile Acids Modified by the Gut Microbiome. *Microorganisms*, 10(1): 68. <https://doi.org/10.3390/microorganisms10010068>
- [72] Heinken A, Ravcheev DA, Baldini F, et al., 2019, Systematic Assessment of Secondary Bile Acid Metabolism in Gut Microbes Reveals Distinct Metabolic Capabilities in Inflammatory Bowel Disease. *Microbiome*, 7(1): 75. <https://doi.org/10.1186/s40168-019-0689-3>
- [73] Mardinoglu A, Agren R, Kampf C, et al., 2013, Integration of Clinical Data with A Genome-Scale Metabolic Model of the Human Adipocyte. *Mol Syst Biol*, 9: 649. <https://doi.org/10.1038/msb.2013.5>

Publisher's note

Bio-Byword Scientific Publishing remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.